

Are Microplastics a Vector for Chemical Contaminants?

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Today's Presentation

- Overview of Microplastics Issue
- Microplastic Survey in Muskegon Lake
- Experimental Results for Persistent Organic Pollutants (POPs) adsorption by microplastics in Muskegon Lake

What are microplastics?

- Plastics that are less than 5 mm in length



Where do microplastics come from?

- Estimated that 8.3 billion metric tons of plastic produced to date.
- 79% of this material deposited in landfills and the natural environment.
- Primary sources of microplastics (e.g., nurdles, additives to consumer products)
- Secondary sources of microplastics as a result from breakdown of larger plastic materials.



Where are we finding microplastics ?

- Air and dust
- Food and beverages
- Cosmetics
- Wastewater
- Industrial wastewater
- Surface water
- Sediments and soil
- Wildlife
- Karst groundwater

- And everywhere else we look



Are humans exposed to microplastics?

- Humans consume over 100 microplastic particles/day and can inhale up to 170/day.
- Bottled water increases microplastic consumption 2 to 3x that of tap water.
- A great deal of variation in reported microplastic data → attributed to lack of standard methods of analysis.
- Need to develop standardized methods for analysis of microplastics.



Are there adverse health effects from exposure to microplastics?



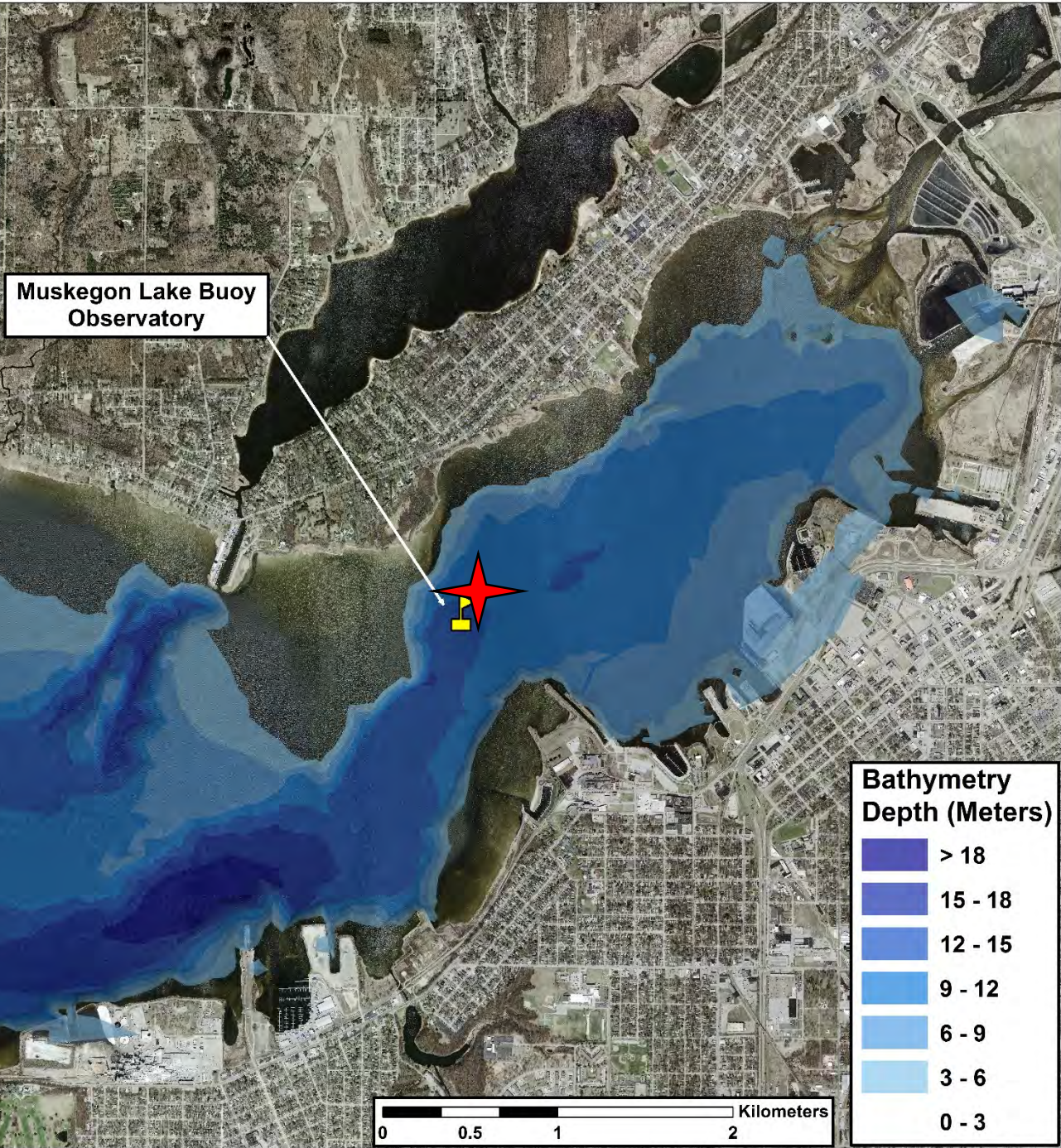
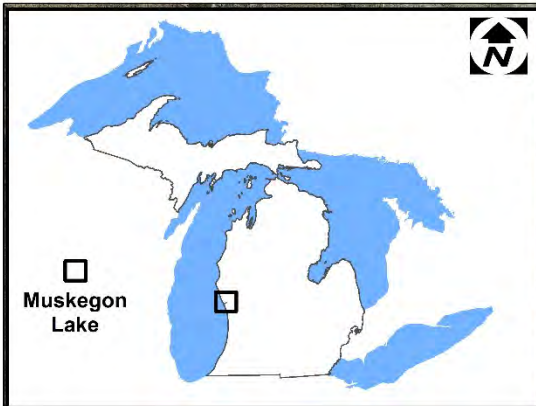
Source: journals.openedition.org

- Adverse effects on wildlife currently under investigation. Some studies show neutral effects, others show negative effects.
- Adverse effects on humans are largely unknown.
- Exposure to heavy metals and additives used in plastic materials.
- Microplastics can concentrate legacy and emerging contaminants from the environment.
- Vectors for pathogens and viruses.

Source: Foley et al. 2018. *Science of the Total Environment* 631 (2018): 550-559.

Microplastics in Muskegon Lake (water column)

- Type of Microplastic
- Color



Methods – Microplastic Survey

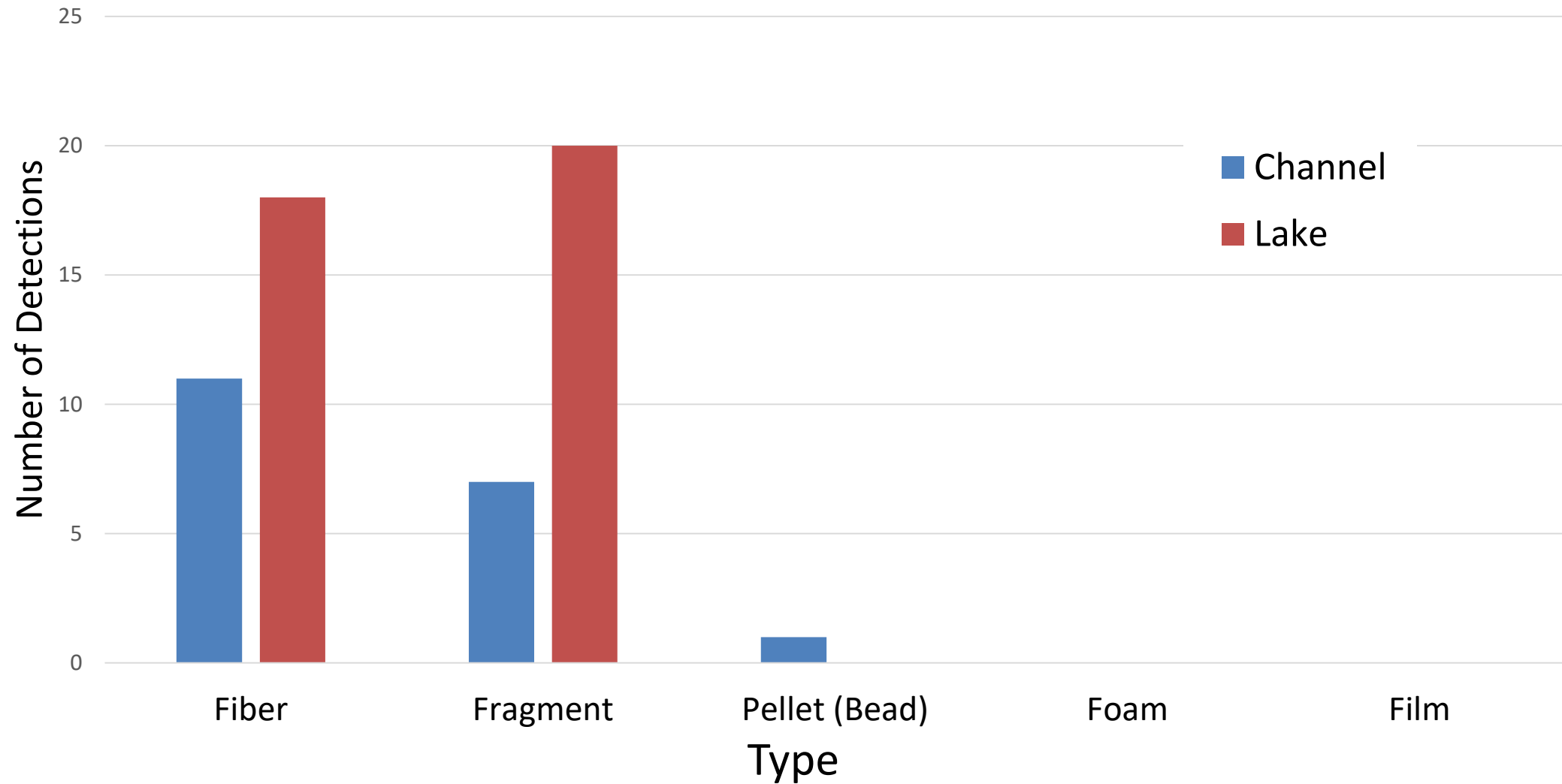
- Grab Samples (Cleaned/Fired Glass Jars)
- 2 sites: channel and buoy
- Modified NOAA protocol
 - Sieving (5 μm), organic digestion, density separation (<1.8 g/L), and isolation on 0.45 μm filter
- Microscopic analysis (Counting and Sizing)
 - Zeiss SteREO Discovery V20 Microscope
 - Size limit of detection 5 μm

Methods – Microplastic Survey Results

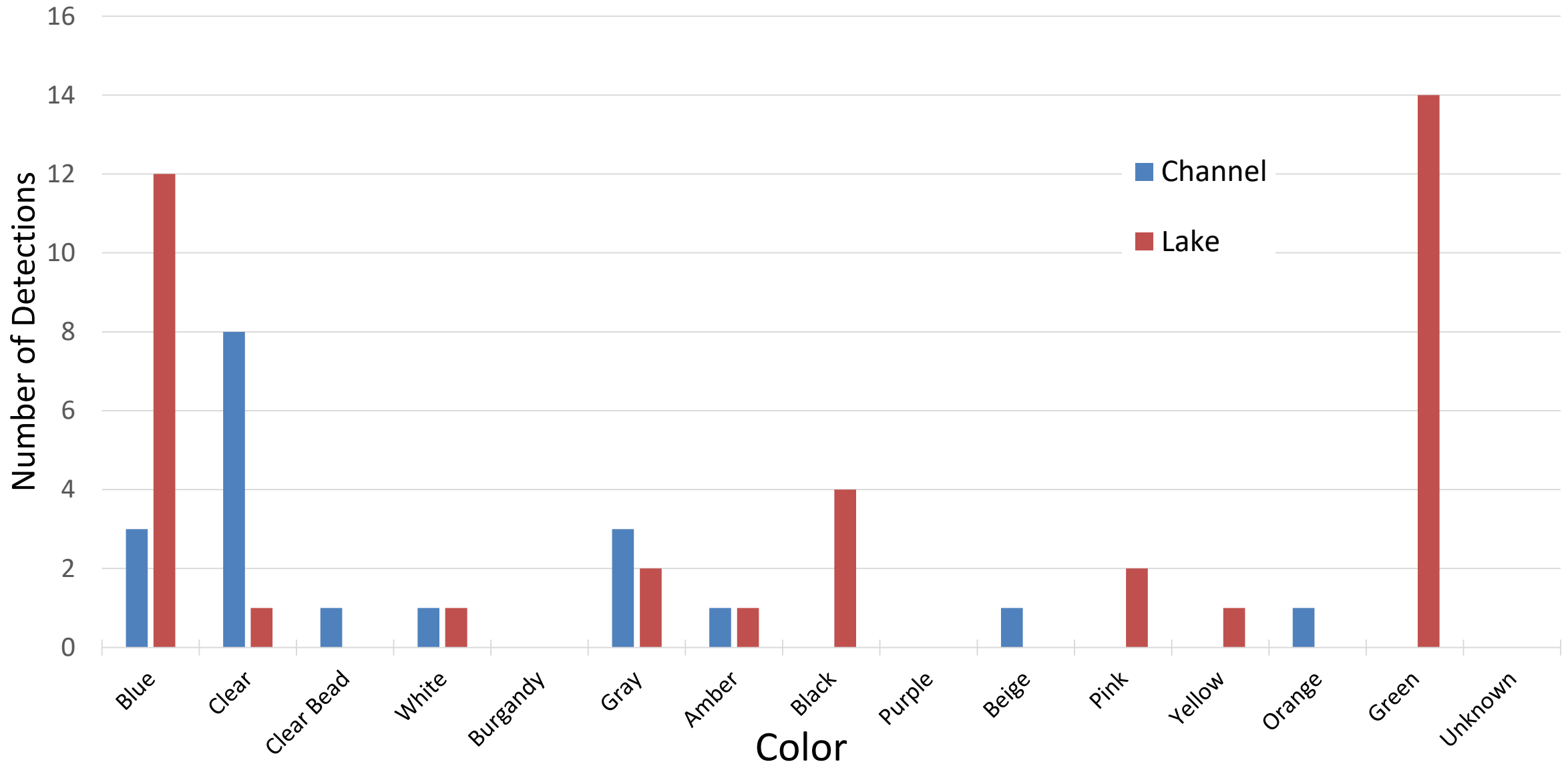
Microplastic Densities:

- Lake open water: 31 particles/L
- Channel: 12 particles/L

Type of Microplastics Detected in Muskegon Lake

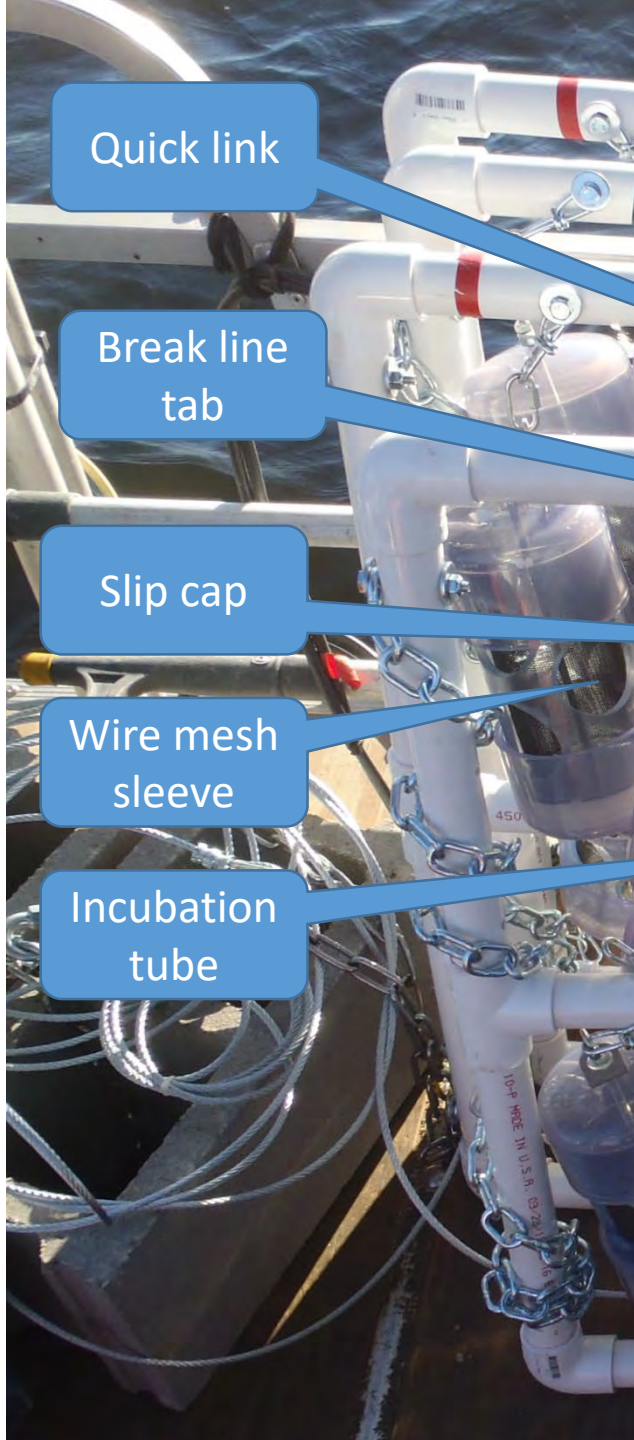


Color Distribution of Microplastics Detected in Muskegon Lake



Experimental Deployment of Microplastics in Muskegon Lake

- Deploy 3 types of Microplastics (polyester; polypropylene; polyethylene)
- 2 sites; 2 depths
- Collect water (Van Dorn bottles) and sediment samples at time of deployment
- Retrieve after 1 and 3 months
- Analyze for variety of contaminants



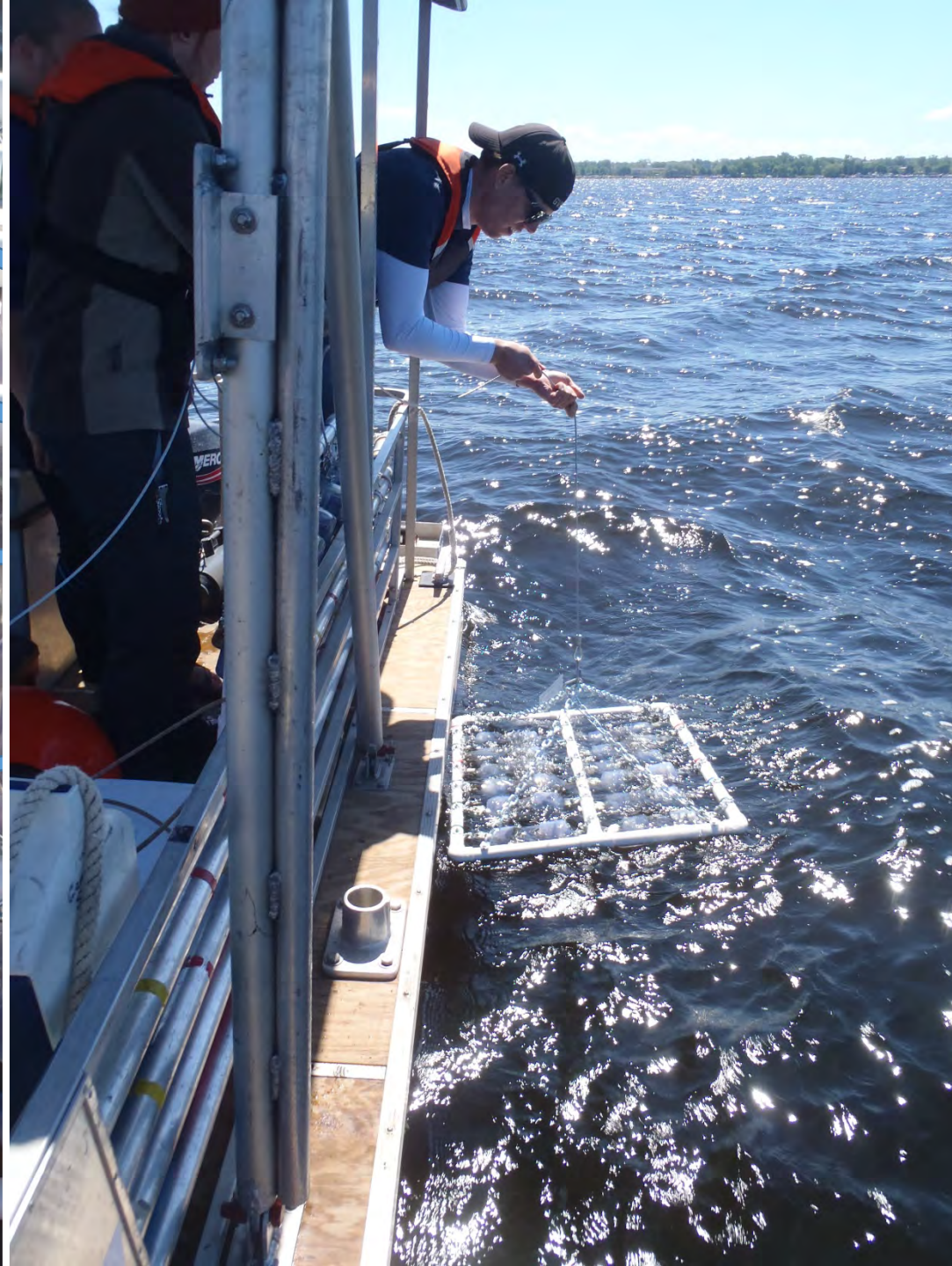
Quick link

Break line
tab

Slip cap

Wire mesh
sleeve

Incubation
tube



View through the water column

Surface Buoy

Muskegon Lake/near bouy
X2 set ups – one for 1 month, one for 3 months

View through the water column

Channel/near Lake Michigan
X2 set ups – one for 1 month, one for 3 months

~2 m

Cable



~2m

Cable

PVC frame

*weighted with chain for sediment deployment roughly ~2' x ~3'

Lake Incubation

Channel Incubation

Mooring weight/cinder blocks

Channel wall

Sediment

Sediment



Target Contaminants

- Polycyclic Aromatic Hydrocarbons (PAHs) – 16 Compounds
- Polychlorinated Biphenyls (PCBs) – 27 Congeners
- Chlorinated Pesticides (OCs) – 12 Compounds
- Polybrominated Biphenyl Ethers (PBDEs) – 9 Congeners
- Perfluoroalkyl substances (PFAS) – 7 Compounds
- Heavy Metals – Cr, Mn, Cu, Zn, As, Se, Ag, Cd, and Pb

Sample Preparation – Water Samples

Aqueous Samples (POPs)

- Liquid-Liquid extraction with dichloromethane
- Extract drying under sodium sulfate
- Silica gel fractionation
- Exchange to hexane and concentrate to 1.0 mL final volume

Sample Preparation – Plastic Samples

Plastics (POPs)

- Accelerated solvent extraction (ASE 300) with hexane
- Silica gel fractionation
- Exchange to hexane and concentrate to 1.0 mL final volume

Plastics (Metals)

- Microwave digest with nitric acid and dilute to final volume 50 mL in DI water

Associated Quality Control

Per Sample – 18 POP Surrogates and 11 POP internal standards

- Per Preparation Batch
- All samples prepared in triplicate
- Reagent blank & reagent blank spike
- Silica gel quality control spike
- Sample matrix spike
- Analytical sample duplicate
- Analytical spike sample

Sample Analysis

PAHs and PCBs

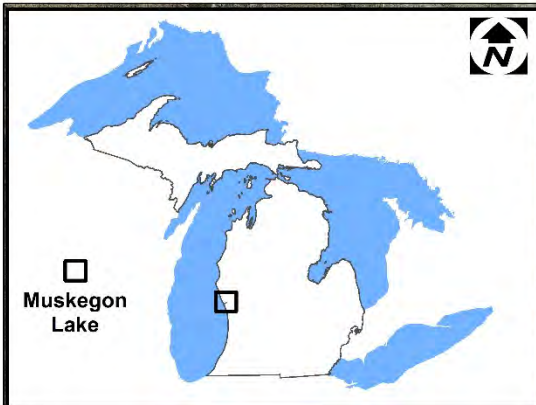
- Shimadzu QP-2010 SE Gas Chromatography Mass Spectrometer

DDT, DDE, DDD, OCs, and PBDEs

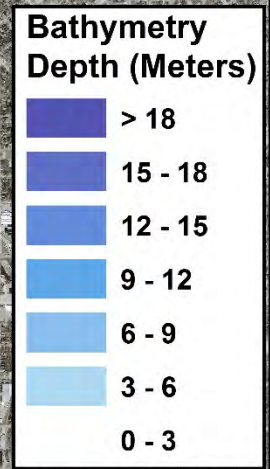
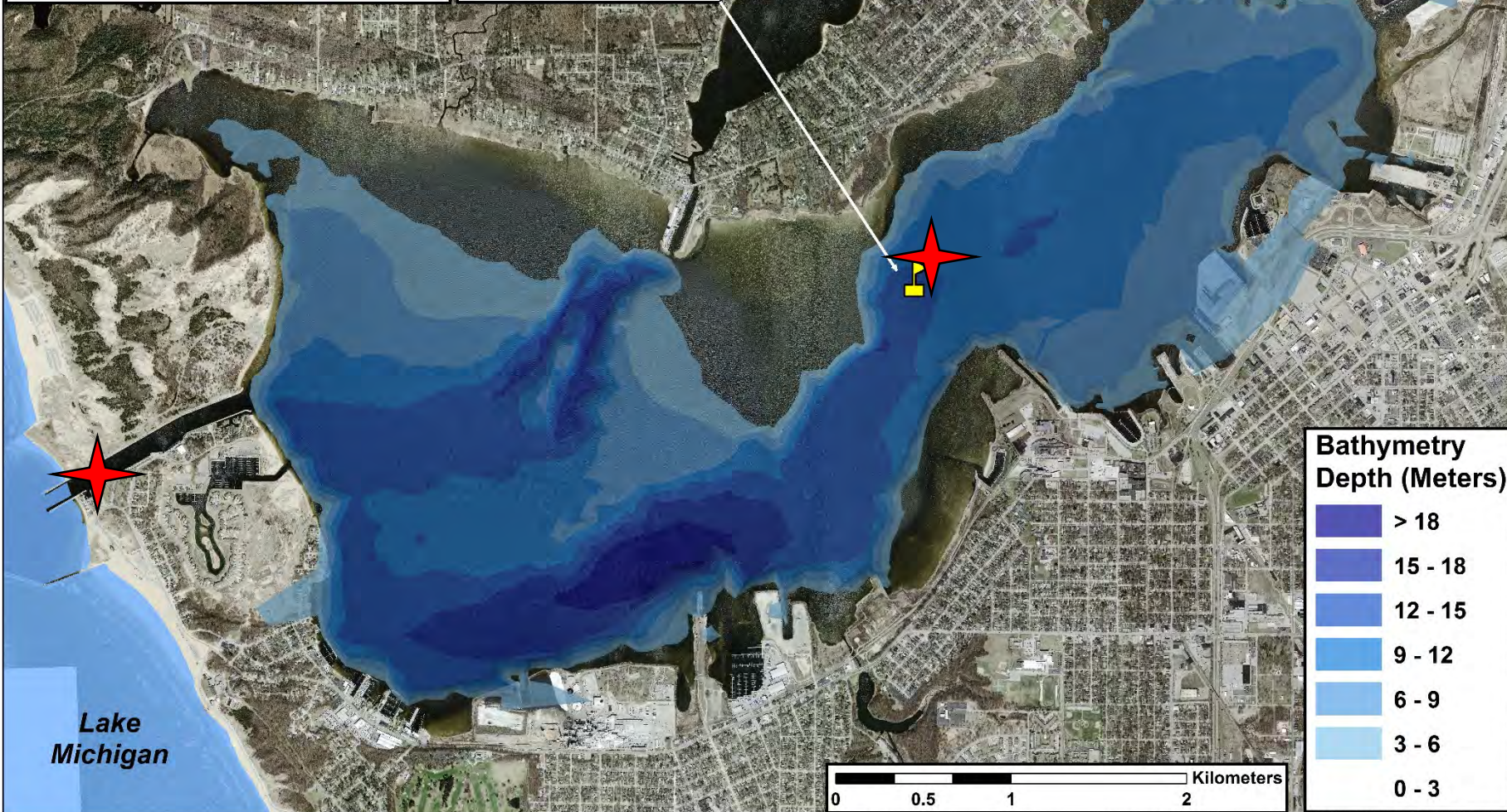
- Agilent 6890 Gas Chromatograph coupled to an Autospec Ultima High Resolution Mass Spectrometer

Metals

- VG PQ ExCell Inductively coupled plasma mass spectrometer



Muskegon Lake Buoy Observatory



Research Questions Addressed

- Does exposure duration influence adsorption of chemical pollutants and biofilm formation?

Virgin polyethylene



Polyethylene,
1-month



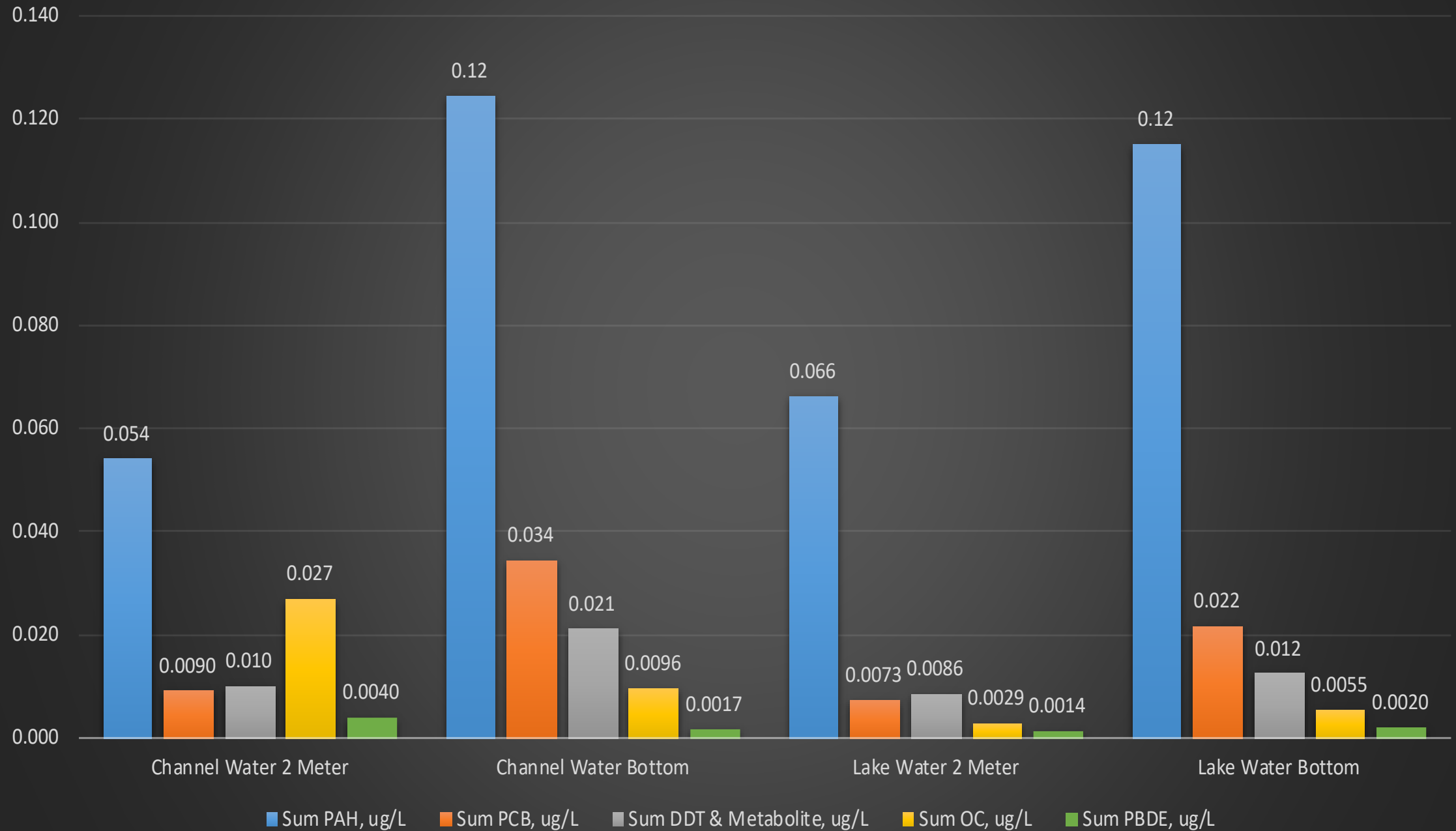
Polyethylene,
3-month



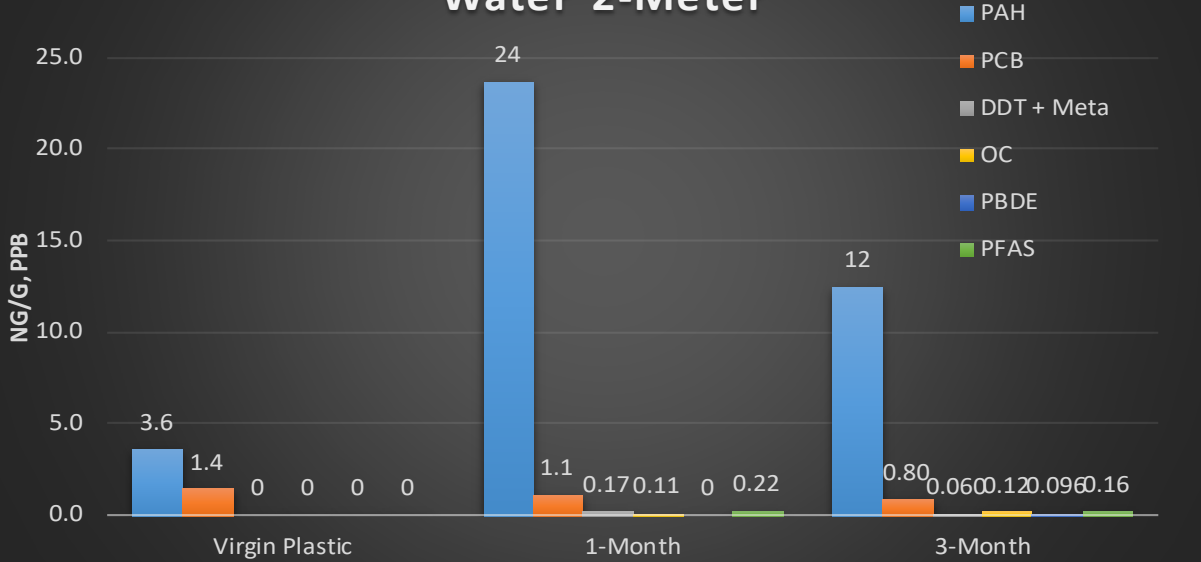
Research Questions Addressed

- Does exposure duration influence adsorption of chemical pollutants and biofilm formation?
- Is plastic type an important factor in accumulation of chemical contaminants and biofilms?
- What role does location play in accumulation of chemical contaminants and biofilms?

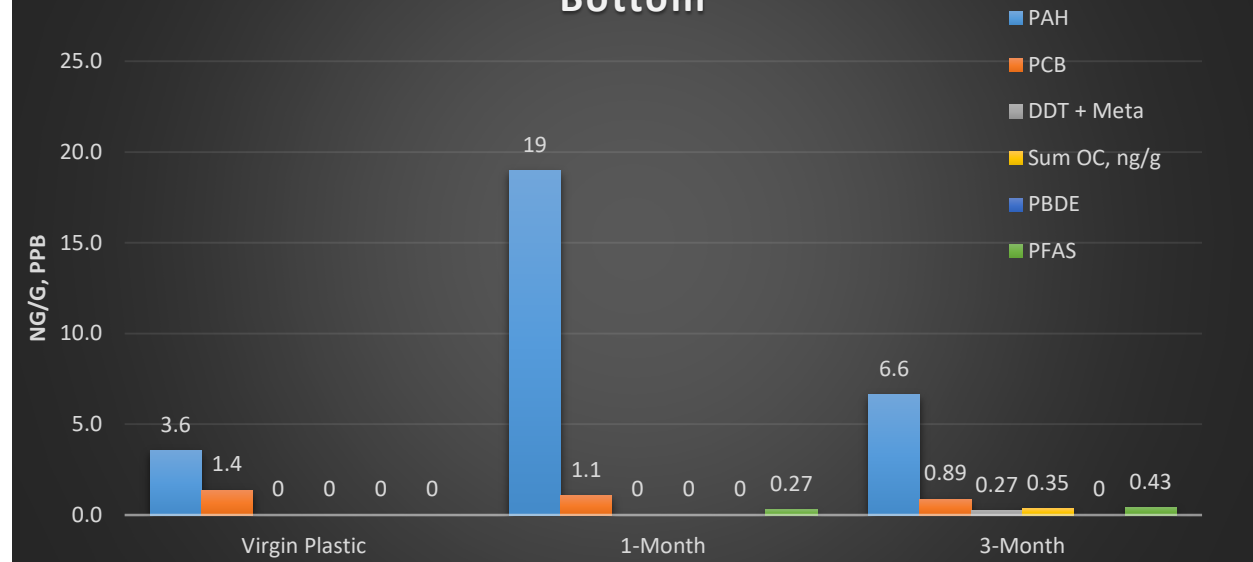
Figure 1: POP Class Sums for Lake Muskegon Aqueous Samples, in $\mu\text{g/L}$



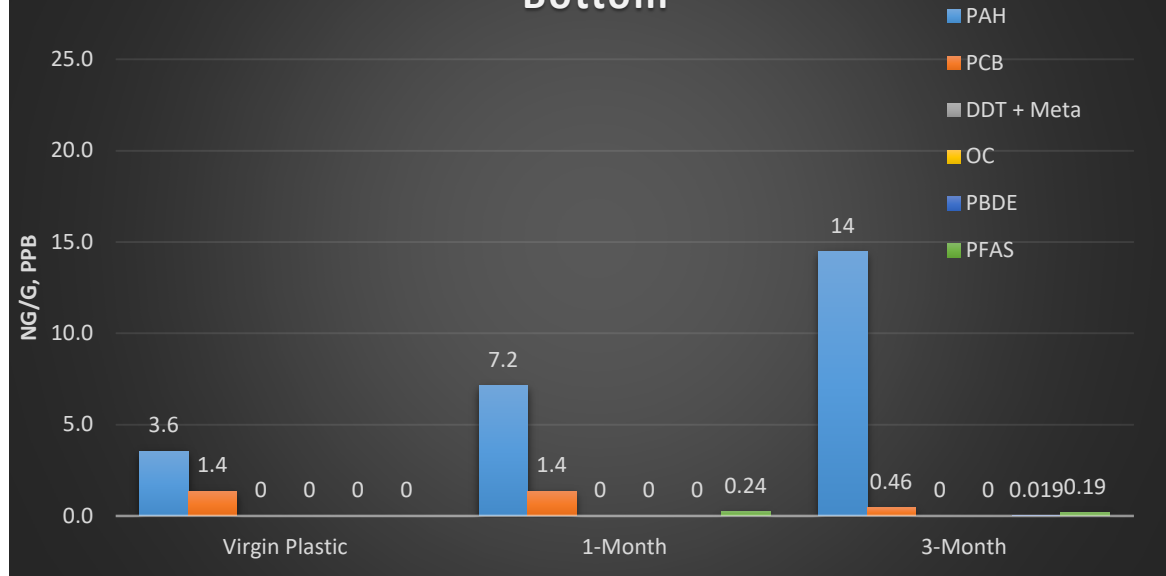
Sum Target POP on Polyester for Channel Water 2-Meter



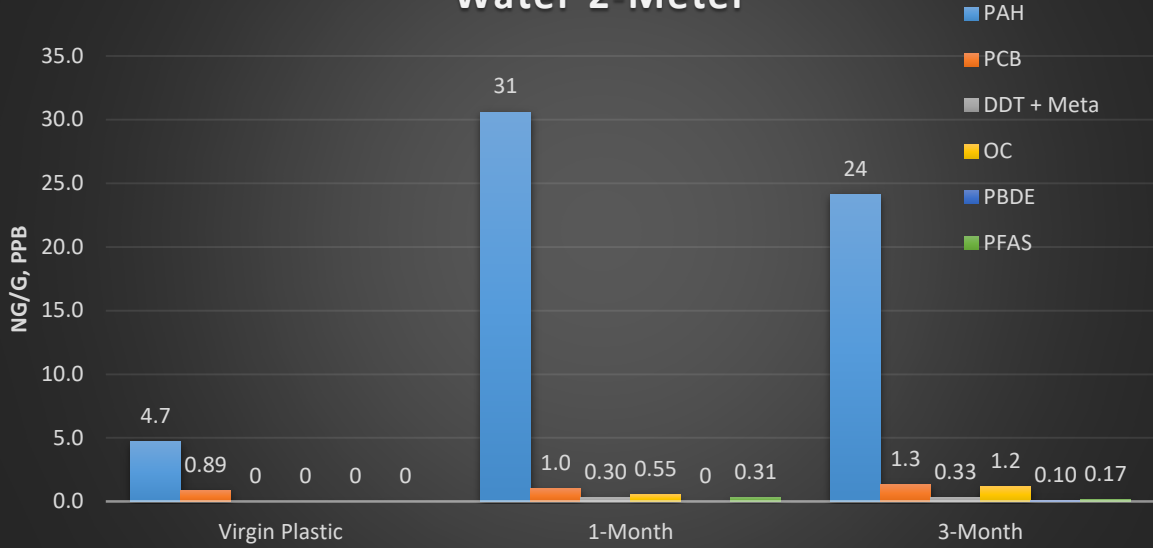
Sum Target POP on Polyester for Channel Water Bottom



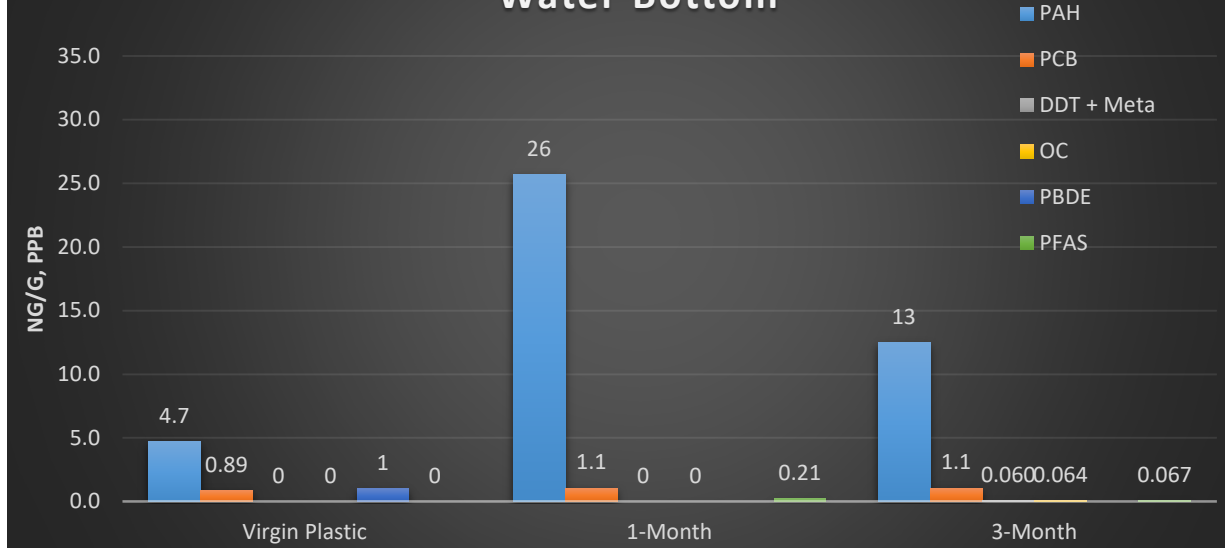
Sum Target POP on Polyester for Lake Water Bottom



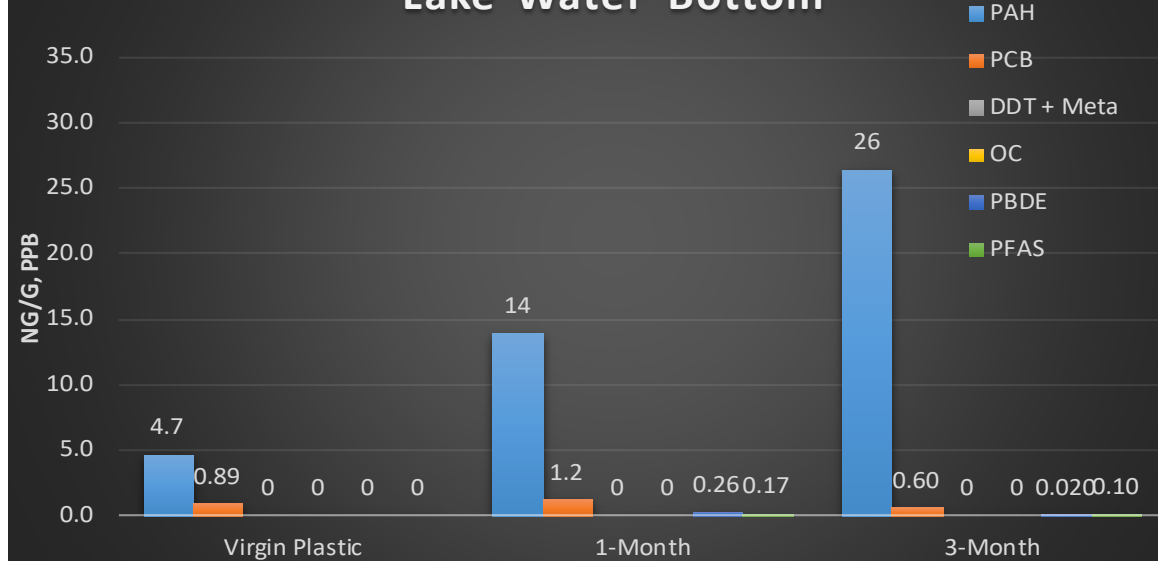
Sum Target POP on Polypropylene for Channel Water 2-Meter



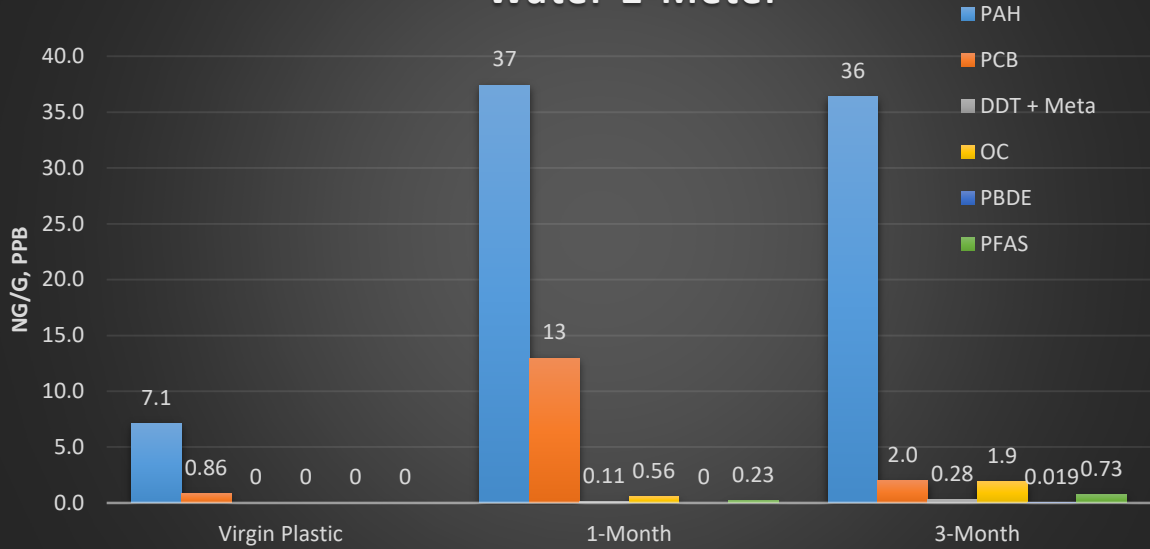
Sum Target POP on Polypropylene for Channel Water Bottom



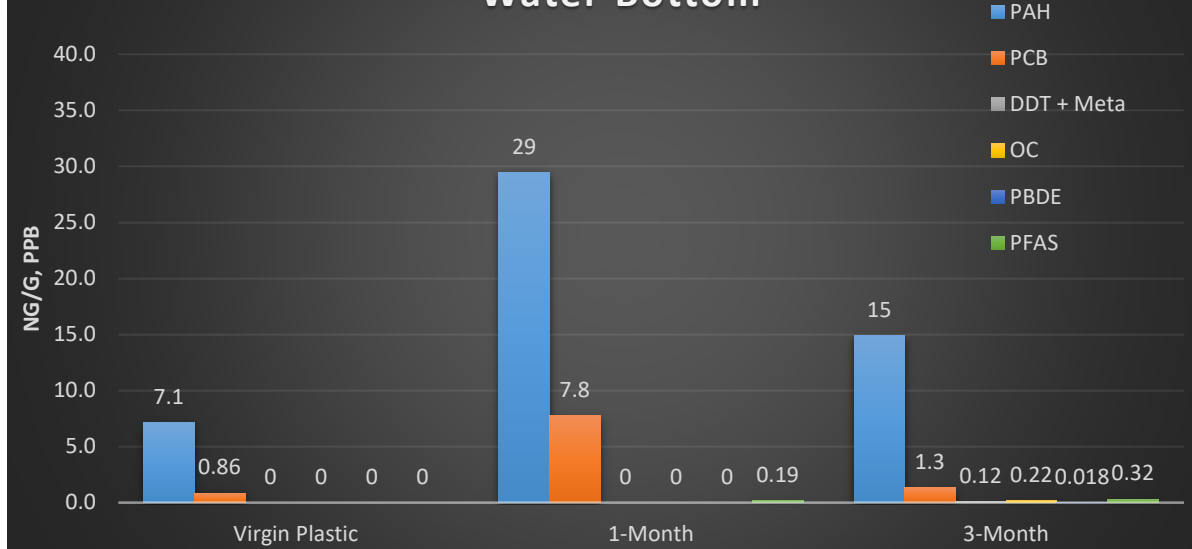
Sum Target POP on Polypropylene for Lake Water Bottom



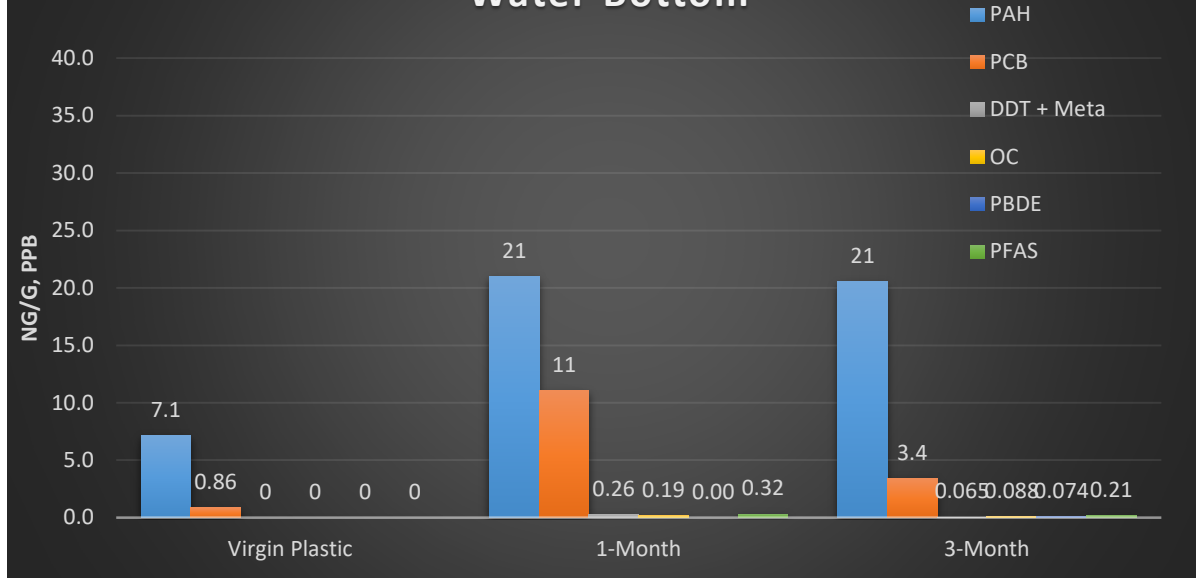
Sum Target POP on Polyethylene for Channel Water 2-Meter



Sum Target POP on Polyethylene for Channel Water Bottom



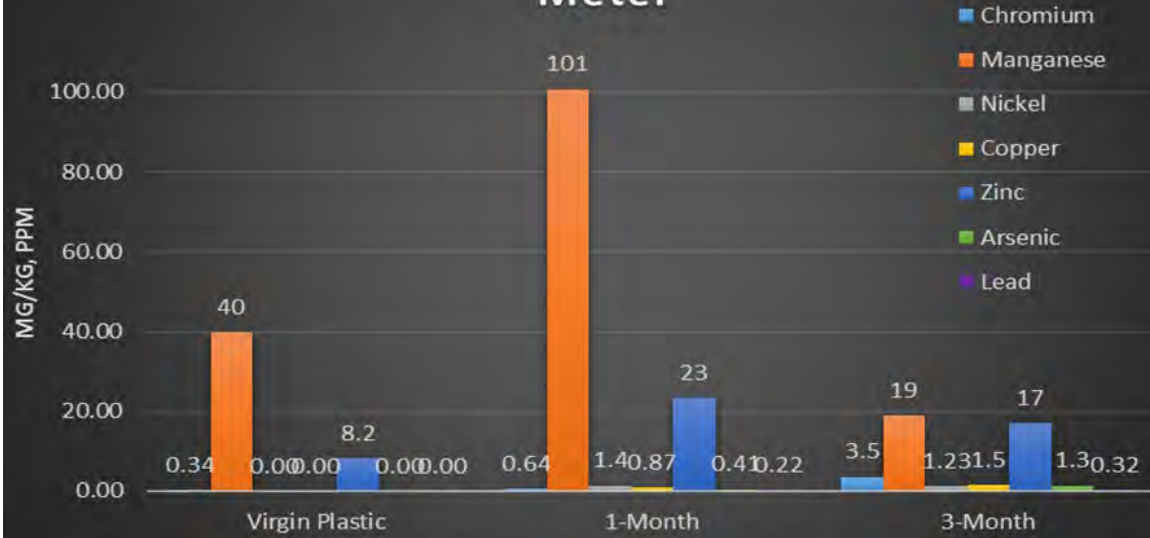
Sum Target POP on Polyethylene for Lake Water Bottom



POP Results

- POP concentrations from highest to lowest: polyethylene > polypropylene > polyester.
- PAHs were the most prevalent POP found on the microplastics. Channel sample concentrations peaked at 1-mo, then declined at 3-mo; lake bottom samples → reverse
- Only slight concentrations of DDT, DDE, DDD, PBDEs, and PFAS were found on plastics after 1-mo and 3-mo deployments.

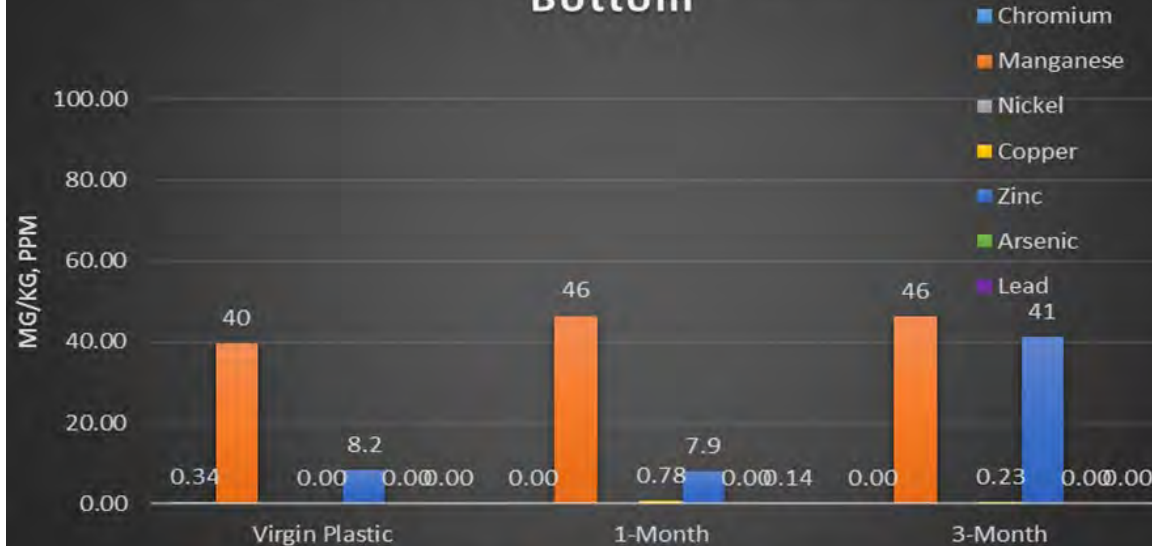
Metals on Polyester for Channel Water 2-Meter



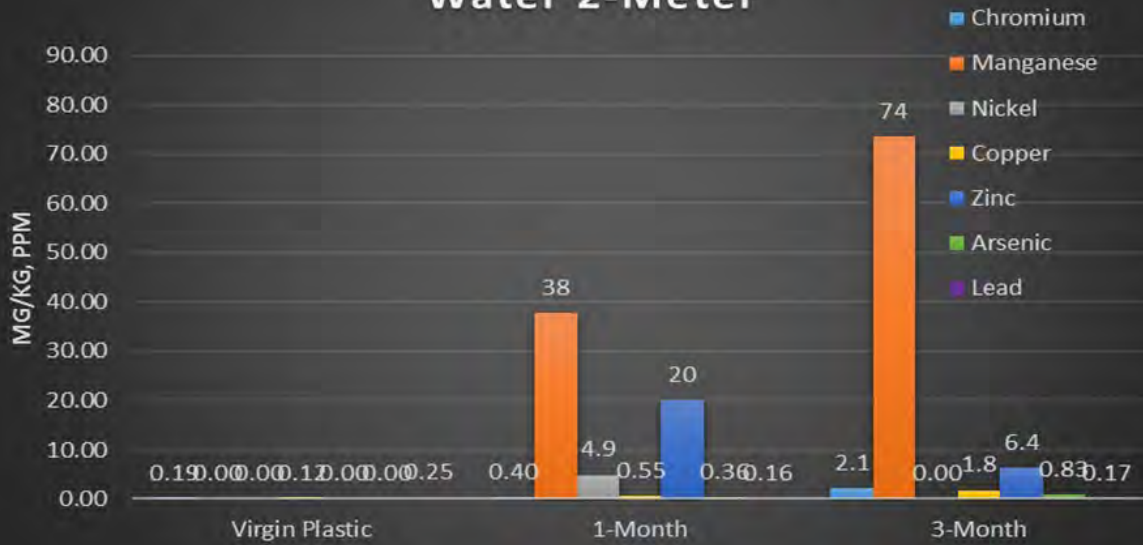
Metals on Polyester for Channel Water Bottom



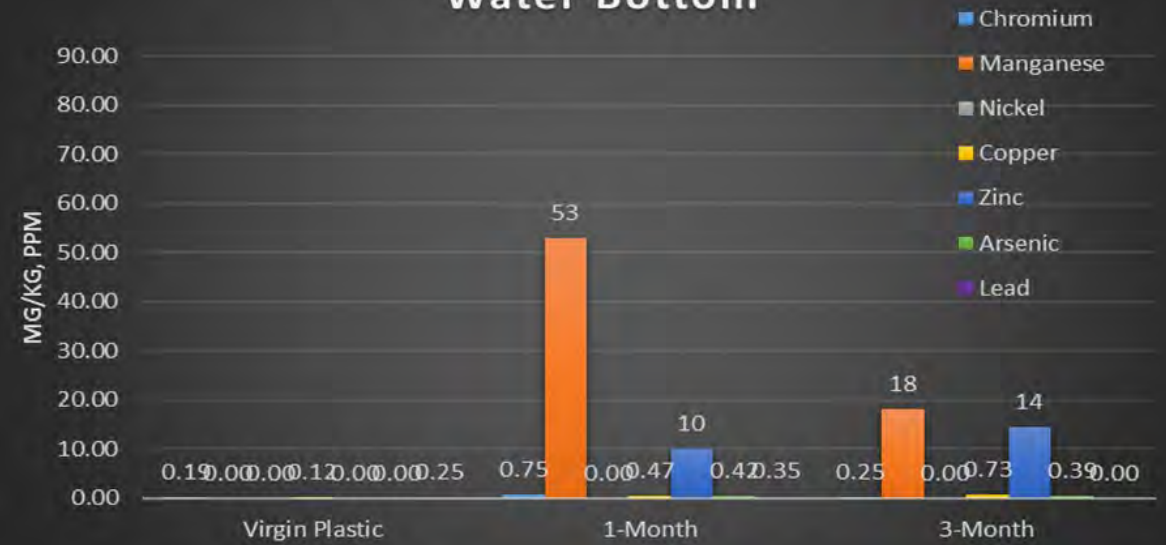
Metals on Polyester for Lake Water Bottom



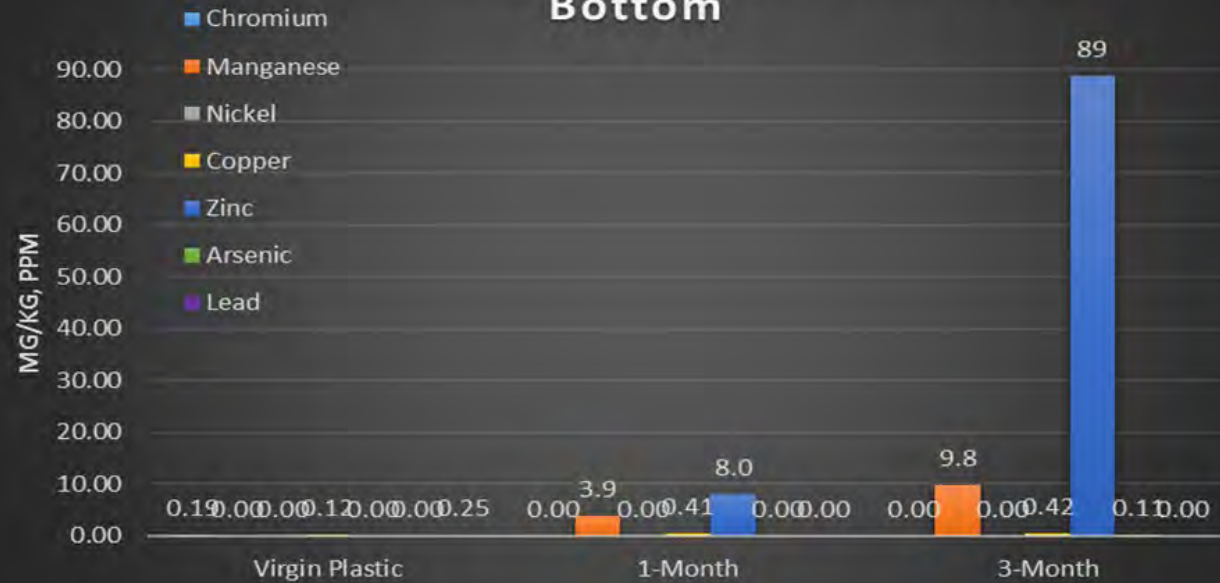
Metals on Polyproylene for Channel Water 2-Meter



Metals on Polyproylene for Channel Water Bottom



Metals on Polyproylene for Lake Water Bottom



Metals on Polyethylene for Channel Water 2-Meter



Metals on Polyethylene for Channel Water Bottom



Metals on Polyethylene for Lake Water Bottom



Metals Results

- Significant concentrations of manganese, and to a lesser degree zinc, were observed on the polyester material before deployment (manufacturing or processing artefact?)
- Mn and Zn were the most abundant metals after deployment.
- Like the POPs, 4 out of 6 of the channel water metals concentrations spiked at 1-month and then declined at 3-months.

PFAS Results

- Most common PFAS's were PFHxA, PFHpA, and PFOA
- Background water sample concentrations low:
 - Channel (surface): 2.8 ng/L (ppt)
 - Lake (surface): 3.3 ng/L (ppt)
- PFAS's were concentrated 24 to 259× background water samples PFAS concentrations very variable, suggesting effect of biofilm

Summary

- Within 1-month, certain microplastics concentrated specific POPs up to $380 \times$ aqueous background concentration.
- Mn and Zn were concentrated at a minimum of 90 to $600 \times$ aqueous background concentrations.
- POP and metals adsorption varied temporally and spatially at the locations of this study.
- PFAS also concentrated but overall impact to fish likely minimal

Next Steps

- Publish findings
- Further work on PFAS
- Feeding experiments:
 - Dreissenid mussels
 - Yellow perch

Acknowledgments

Sampler Design

- Maggie Oudsema (AWRI)

Sampler Fabrication, Deployment, and Retrieval

- Emily Kindervater, Rachel Orzechowski, Paige Kleindl, & Mike Hassett (AWRI)

Plastic Preparations, Sample Preparation & Analysis

- Jessica Porter & Kathryn Gunderson (ISTC)

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- BiRirmingham Illinois Partnership for Discovery, EnGagement, and Education (BRIDGE)
- Illinois Indiana Sea Grant (NA18OAR4170082)

Questions?



Extra Slides

Sample Preparation – Water Samples

Aqueous Samples (POPs except PFAS)

- Liquid-Liquid extraction with dichloromethane
- Extract drying under sodium sulfate
- Silica gel fractionation
- Exchange to hexane and concentrate to 1.0 mL final volume

Aqueous Samples (PFAS) per US EPA Method 537

- Solid Phase Extraction (SPE) with Agilent Bond Elut-LMS
- Blow to dryness under nitrogen
- Reconstituted to 1.0 ml in methanol-water (96:4)

Sample Preparations

Plastics (POPs except PFAS)

- Accelerated solvent extraction (ASE 300) with hexane
- Silica gel fractionation
- Exchange to hexane and concentrate to 1.0 mL final volume

Plastics (PFAS)

- Solid-Liquid extraction with methanol and centrifugation
- Exchange to 60% ammonia acetate (20mM) : 40% methanol and concentrate to 1.0 mL final volume

Plastics (Metals)

- Microwave digest with nitric acid and dilute to final volume 50 mL in DI water

Associated Quality Control

Per Sample – 18 POP Surrogates and 11 POP internal standards

For PFAS – 7 Isotope PFAS Surrogates and 2 internal standards

- Per Preparation Batch
- All samples prepared in triplicate (except PFAS in duplicates)
- Reagent blank & reagent blank spike
- Silica gel quality control spike (except PFAS)
- Sample matrix spike
- Analytical sample duplicate
- Analytical spike sample

Sample Analysis

PAHs and PCBs

- Shimadzu QP-2010 SE Gas Chromatography Mass Spectrometer

DDT, DDE, DDD, OCs, and PBDEs

- Agilent 6890 Gas Chromatograph coupled to an Autospec Ultima High Resolution Mass Spectrometer

PFAS

- Waters Alliance 2695 coupled to a Quattro Micro tandem mass spectrometer

Metals

- VG PQ ExCell Inductively coupled plasma mass spectrometer

PFAS Results

- McNeish et al. (2018) found mean of 13 microplastic particles per fish in Muskegon River
- This study showed a PFAS concentration of 0.87 ng/g (worst case)
 - Assuming a 1.5 mm microfiber mass of $1.5 \times 10^{-5} \text{ cm}^3$:

$$1.5 \times 10^{-5} \text{ cm}^3 \text{ (g/particle)} \times 13 \text{ particles} \times 0.87 \text{ ng/g} = 0.0002 \text{ ng}$$

- Unlikely to have negative effect on fish (but need info on feeding; interactions with POPs; commercial microplastics)

Size Distribution of Microplastics Detected in Muskegon Lake

